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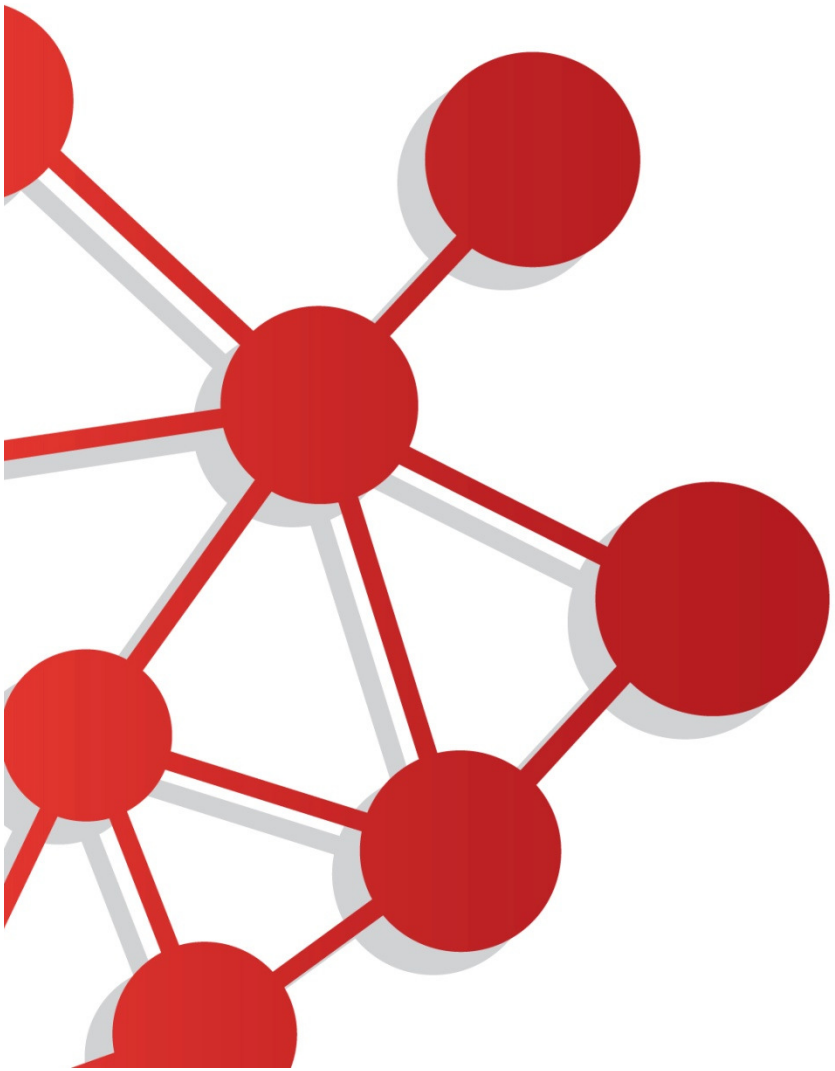

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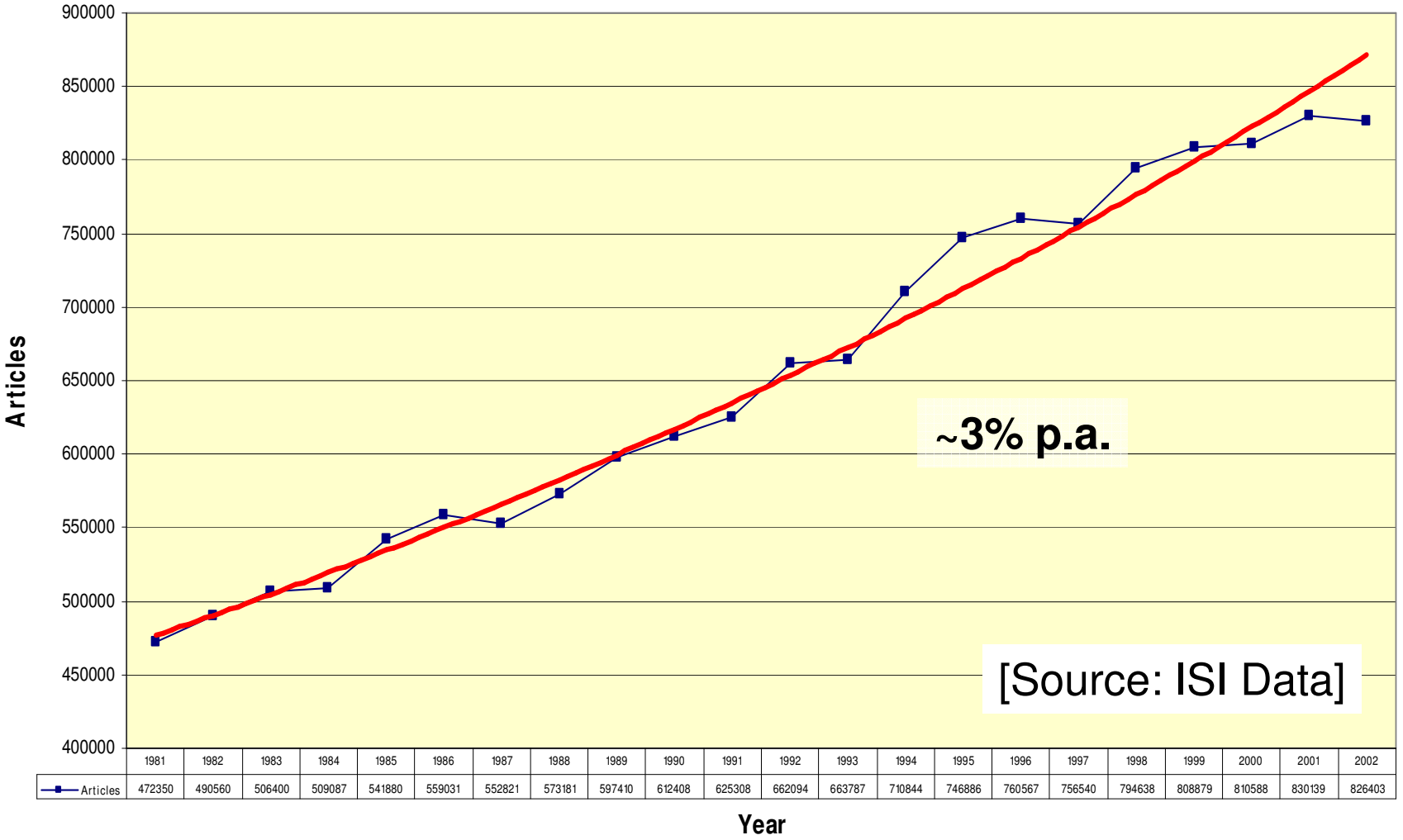

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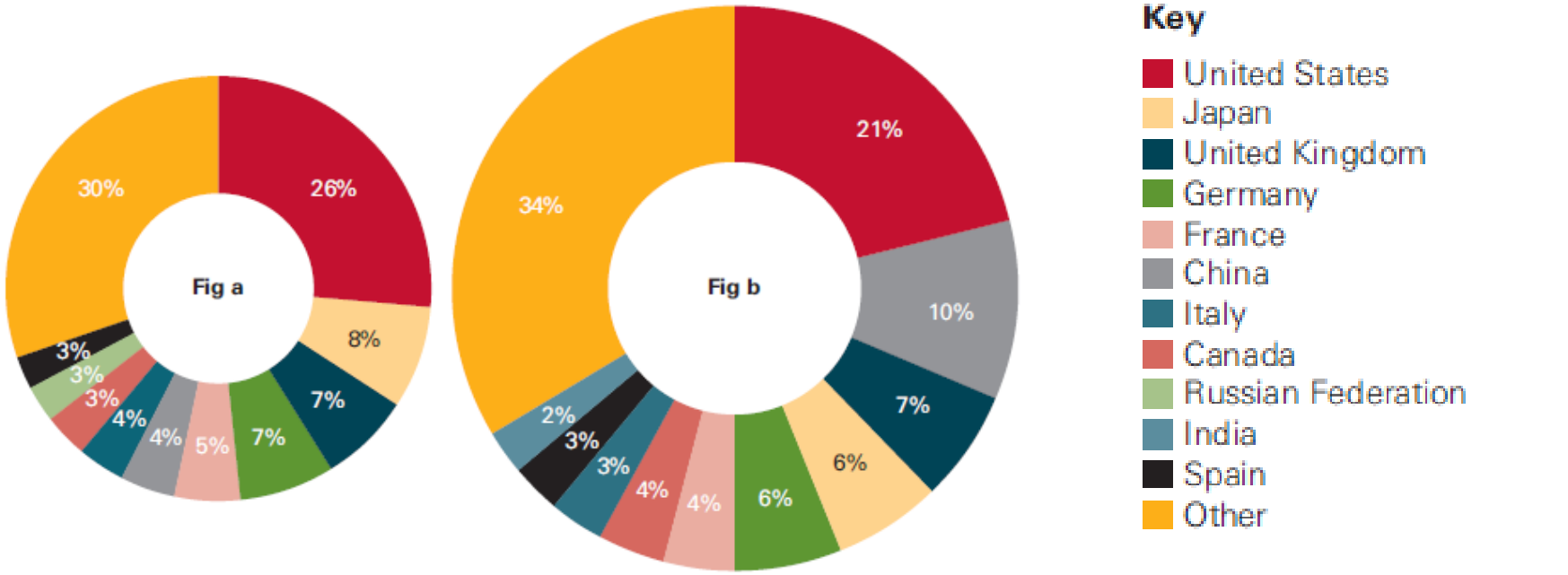

Knowledge, networks, nations: Global scientific collaboration in the 21st century.
RS policy document, March 2011 – data from Scopus

DNA structure - 1953

No. 4356 April 25, 1953 NATURE 737

equipment, and to Dr. G. E. R. Deacon and the captain and officers of R.R.S. *Discovery II* for their part in making the observations.

1 Young, F. B., Gerrard, H., and Jevons, W., *Phil. Mag.*, **40**, 149 (1920).
2 Longuet-Higgins, M. S., *Mon. Not. Roy. Astro. Soc., Geophys. Supp.*, **5**, 285 (1949).
3 Von Arx, W. S., Woods Hole Papers in Phys. Oceanog. Meteor., **11** (3) (1950).
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MOLECULAR STRUCTURE OF NUCLEIC ACIDS

A Structure for Deoxyribose Nucleic Acid

WE wish to suggest a structure for the salt of deoxyribose nucleic acid (D.N.A.). This structure has novel features which are of considerable biological interest.

A structure for nucleic acid has already been proposed by Pauling and Corey[1]. They kindly made their manuscript available to us in advance of publication. Their model consists of three intertwined chains, with the phosphates near the fibre axis, and the bases on the outside. In our opinion, this structure is unsatisfactory for two reasons: (1) We believe that the material which gives the X-ray diagrams is the salt, not the free acid. Without the acidic hydrogen atoms it is not clear what forces would hold the structure together, especially as the negatively charged phosphates near the axis will repel each other. (2) Some of the van der Waals distances appear to be too small.

Another three-chain structure has also been suggested by Fraser (in the press). In his model the phosphates are on the outside and the bases on the inside, linked together by hydrogen bonds. This structure as described is rather ill-defined, and for this reason we shall not comment on it.

This figure is purely diagrammatic. The two ribbons symbolize the two phosphate–sugar chains, and the horizontal rods the pairs of bases holding the chains together. The vertical line marks the fibre axis

We wish to put forward a radically different structure for the salt of deoxyribose nucleic acid. This structure has two helical chains each coiled round the same axis (see diagram). We have made the usual chemical assumptions, namely, that each chain consists of phosphate diester groups joining β-D-deoxyribofuranose residues with 3′,5′ linkages. The two chains (but not their bases) are related by a dyad perpendicular to the fibre axis. Both chains follow right-handed helices, but owing to the dyad the sequences of the atoms in the two chains run in opposite directions. Each chain loosely resembles Furberg's[2] model No. 1; that is, the bases are on the inside of the helix and the phosphates on the outside. The configuration of the sugar and the atoms near it is close to Furberg's 'standard configuration', the sugar being roughly perpendicular to the attached base. There is a residue on each chain every 3·4 A. in the z-direction. We have assumed an angle of 36° between adjacent residues in the same chain, so that the structure repeats after 10 residues on each chain, that is, after 34 A. The distance of a phosphorus atom from the fibre axis is 10 A. As the phosphates are on the outside, cations have easy access to them.

The structure is an open one, and its water content is rather high. At lower water contents we would expect the bases to tilt so that the structure could become more compact.

The novel feature of the structure is the manner in which the two chains are held together by the purine and pyrimidine bases. The planes of the bases are perpendicular to the fibre axis. They are joined together in pairs, a single base from one chain being hydrogen-bonded to a single base from the other chain, so that the two lie side by side with identical z-co-ordinates. One of the pair must be a purine and the other a pyrimidine for bonding to occur. The hydrogen bonds are made as follows: purine position 1 to pyrimidine position 1; purine position 6 to pyrimidine position 6.

If it is assumed that the bases only occur in the structure in the most plausible tautomeric forms (that is, with the keto rather than the enol configurations) it is found that only specific pairs of bases can bond together. These pairs are: adenine (purine) with thymine (pyrimidine), and guanine (purine) with cytosine (pyrimidine).

In other words, if an adenine forms one member of a pair, on either chain, then on these assumptions the other member must be thymine; similarly for guanine and cytosine. The sequence of bases on a single chain does not appear to be restricted in any way. However, if only specific pairs of bases can be formed, it follows that if the sequence of bases on one chain is given, then the sequence on the other chain is automatically determined.

It has been found experimentally[3,4] that the ratio of the amounts of adenine to thymine, and the ratio of guanine to cytosine, are always very close to unity for deoxyribose nucleic acid.

It is probably impossible to build this structure with a ribose sugar in place of the deoxyribose, as the extra oxygen atom would make too close a van der Waals contact.

The previously published X-ray data[2,3] on deoxyribose nucleic acid are insufficient for a rigorous test of our structure. So far as we can tell, it is roughly compatible with the experimental data, but it must be regarded as unproved until it has been checked against more exact results. Some of these are given in the following communications. We were not aware of the details of the results presented there when we devised our structure, which rests mainly though not entirely on published experimental data and stereochemical arguments.

It has not escaped our notice that the specific pairing we have postulated immediately suggests a possible copying mechanism for the genetic material.

Full details of the structure, including the conditions assumed in building it, together with a set of co-ordinates for the atoms, will be published elsewhere.

We are much indebted to Dr. Jerry Donohue for constant advice and criticism, especially on interatomic distances. We have also been stimulated by a knowledge of the general nature of the unpublished experimental results and ideas of Dr. M. H. F. Wilkins, Dr. R. E. Franklin and their co-workers at

- 1 page
- 2 authors
- 1 figure
- no data

The human genome - 2001

- 62 pages, 49 figures, 27 tables

articles

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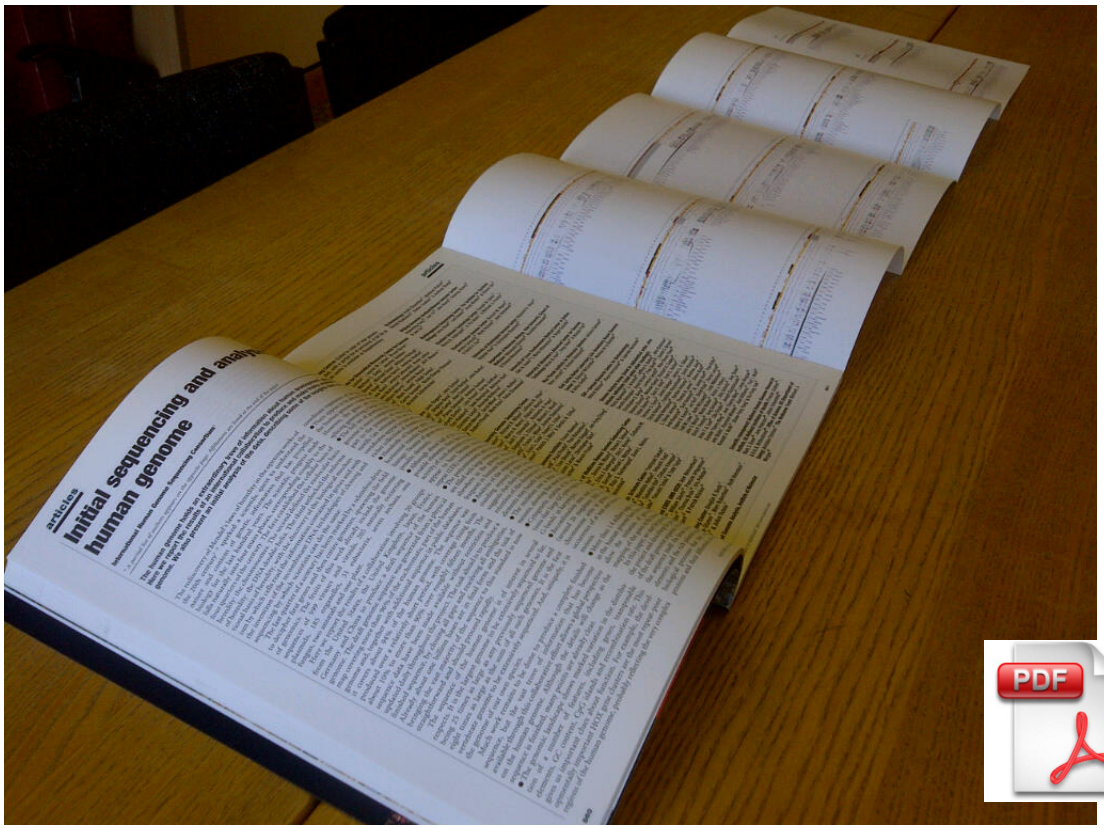

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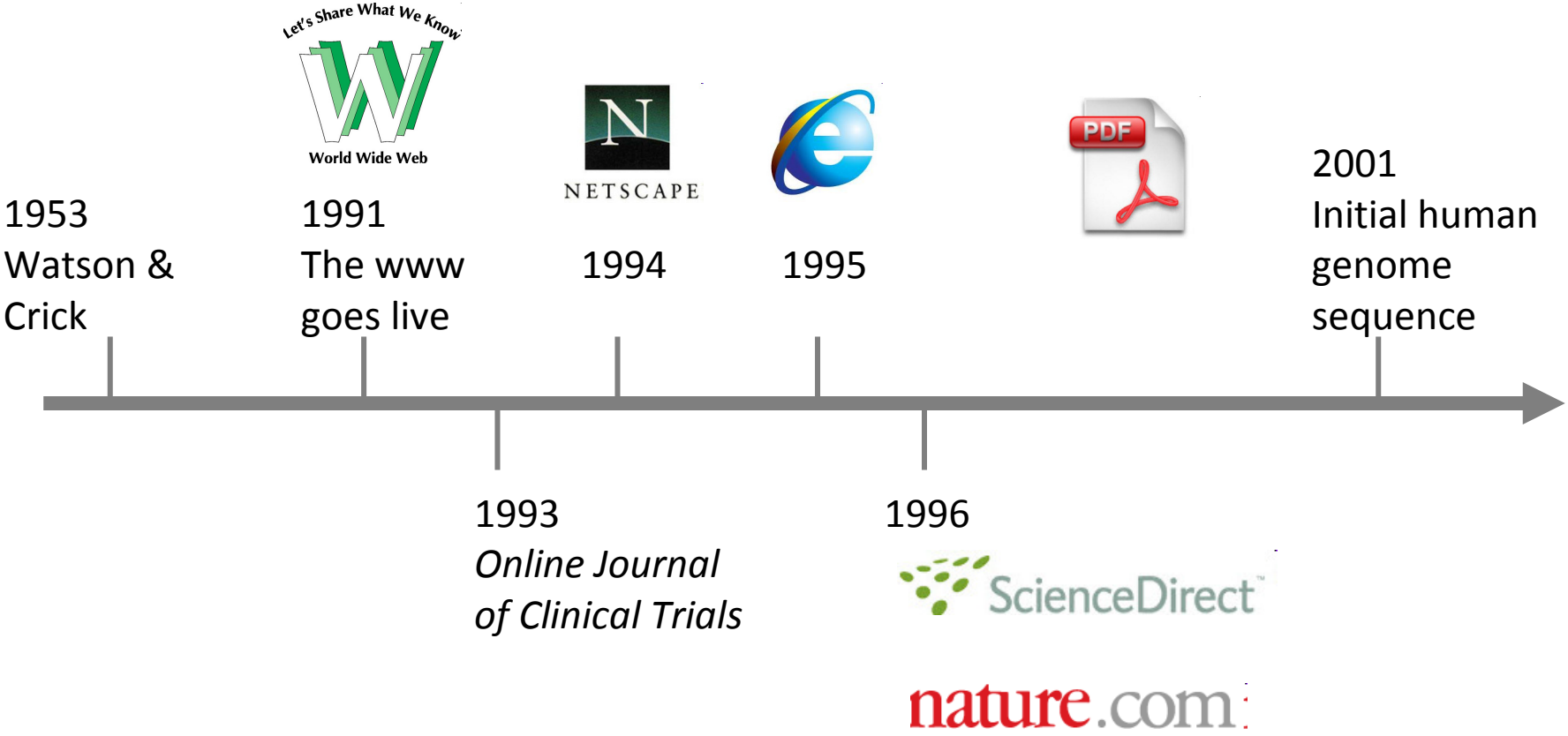

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  - Preprint servers in physical sciences
- Journals are key players in enforcing data deposition
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- Mandates are not enough
  - Challenges in enforcing the MIAME mandate
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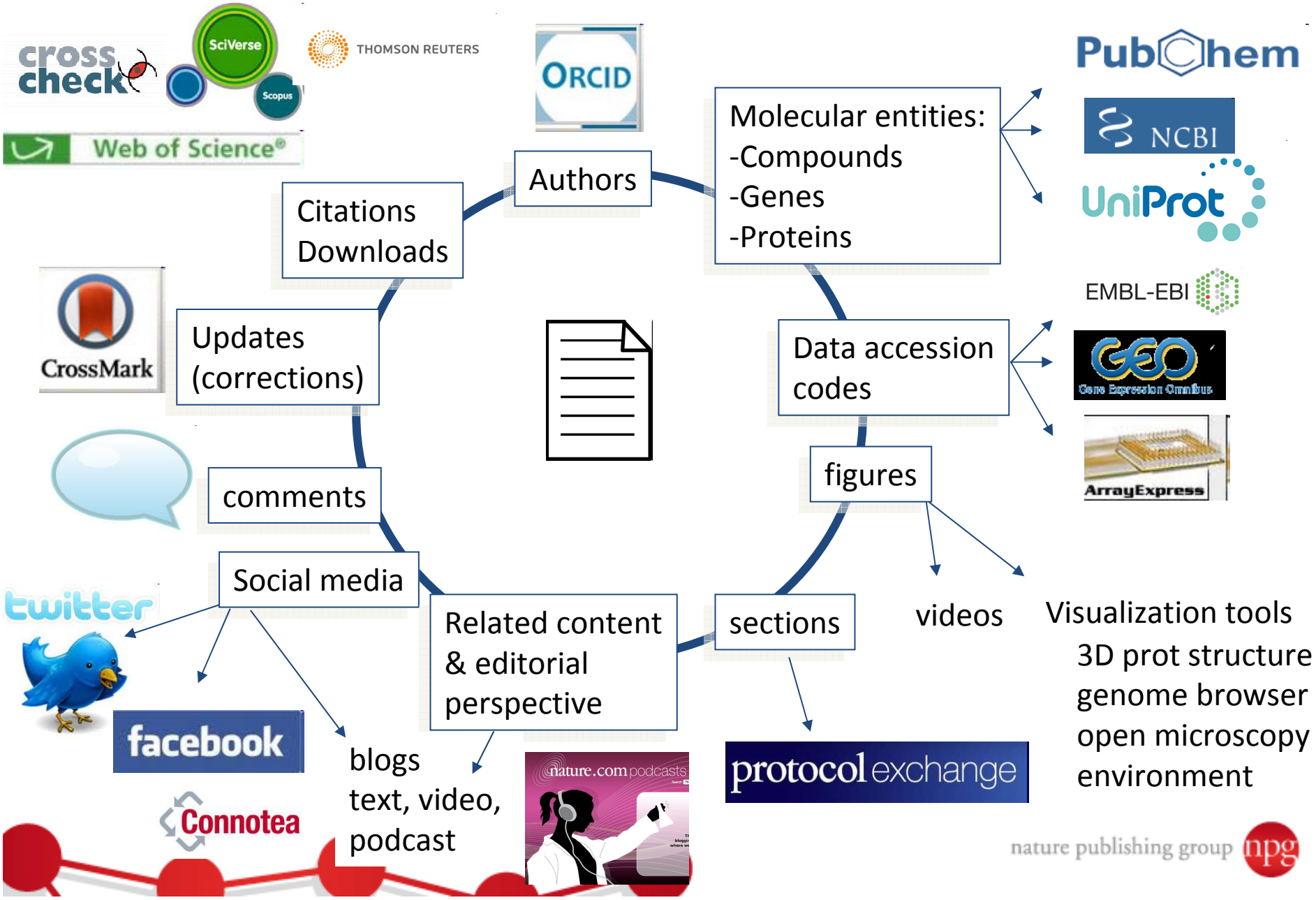

NATURE CHEMICAL BIOLOGY | ARTICLE

Acetylation regulates Cyclophilin A catalysis, immunosuppression and HIV isomerization

Michael Lammers, Heinz Neumann, Jason W Chin & Leo C James

Affiliations | Contributions | Corresponding authors

Nature Chemical Biology **6**, 331–337 (2010) | doi:10.1038/nchembio.342
Received 27 October 2009 | Accepted 14 January 2010 | Published online 04 April 2010

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Abstract

Abstract • Introduction • Results • Discussion • Methods • Additional information • Accession codes • References
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Cyclophilin A (CypA) is a ubiquitous *cis–trans* prolyl isomerase with key roles in immunity and viral infection. CypA suppresses T-cell activation through cyclosporine complexation and is required for effective HIV-1 replication in host cells. We show that CypA is acetylated in diverse human cell lines and use a synthetically evolved acetyllysyl-tRNA synthetase/$tRNA_{CUA}$ pair to produce recombinant acetylated CypA in *Escherichia coli*. We determined atomic-resolution structures of acetylated CypA and its complexes with cyclosporine and HIV-1 capsid. Acetylation markedly inhibited CypA catalysis of *cis* to *trans* isomerization and stabilized *cis* rather than *trans* forms of the HIV-1 capsid. Furthermore, CypA acetylation antagonized the immunosuppressive effects of cyclosporine by inhibiting the sequential steps of cyclosporine binding and

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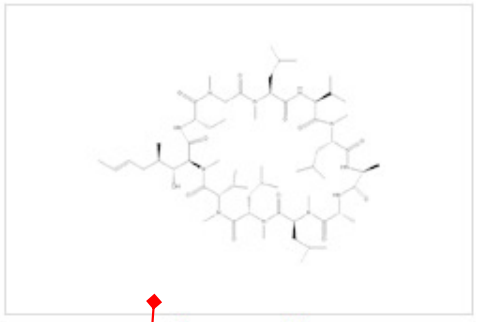

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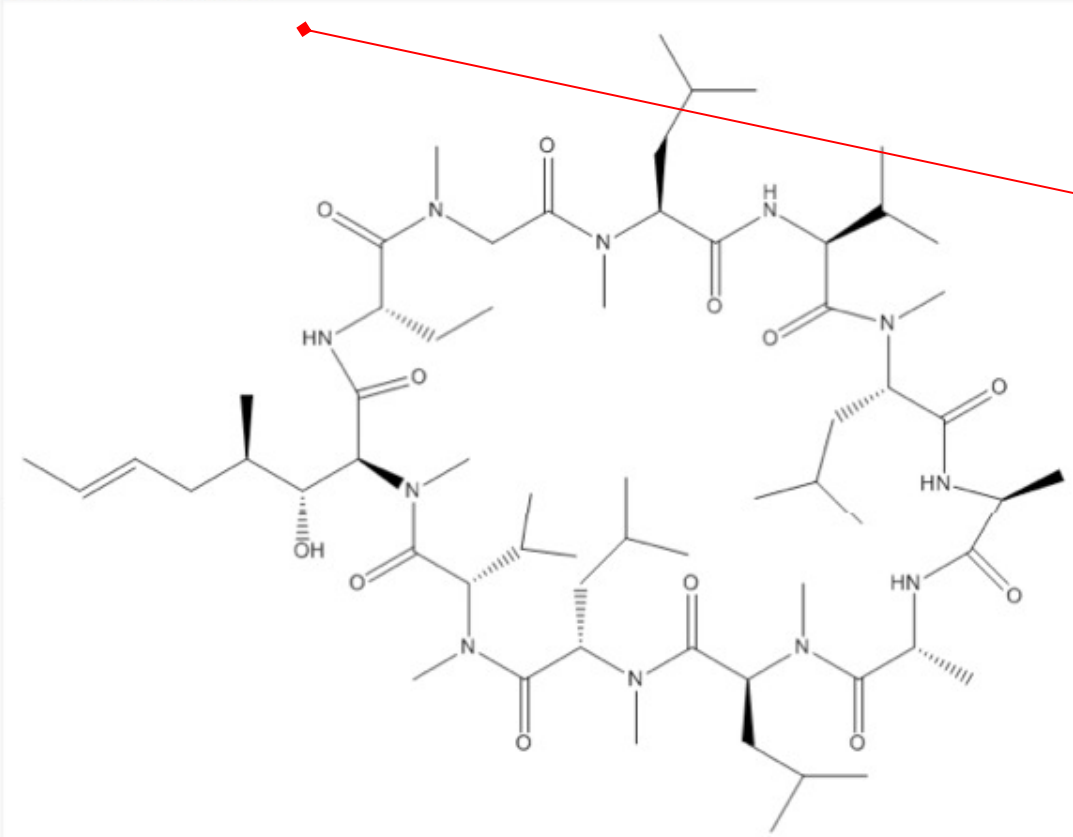

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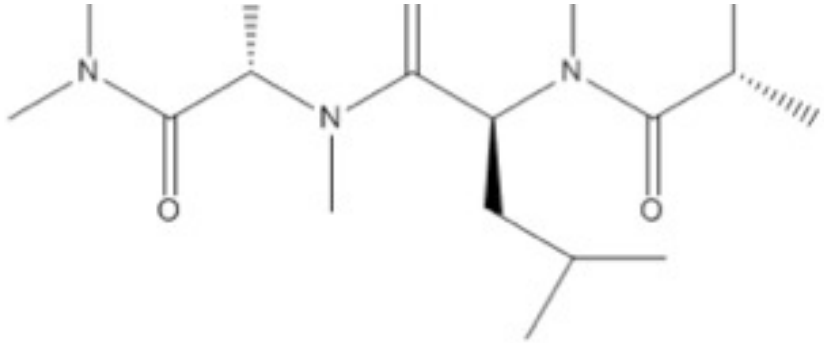

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Synonyms

Cyclosporine
Ciclosporin
Neoral
Cyclosporine (USP)
Antibiotic S 7481F1
BMT-ABA-SAR-MLE-VAL-MLE-ALA-ALA-MLE-MLE-MVA
S-Neoral
Cipol N
HSDB 6881
S 7481F1
DRG 0275
Optimmune
(3S,6S,9S,12R,15S,18S,21S,24S,30S,33S)-30-Ethyl-33-[(1R,2R,4E)-1-hydroxy-2-methylhex-4-en-1-yl]-6,9,18,24-tetraisobutyl-3,21-diisopro
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1,4,7,10,12,15,19,25,28-nonamethyl-3,21-bis(1-m
ethylethyl)-6,9,18,24-tetrakis(2-methylpropyl)-, (3S,6S,9S,12R,15S,18S,21S,24S,30S,33S)-

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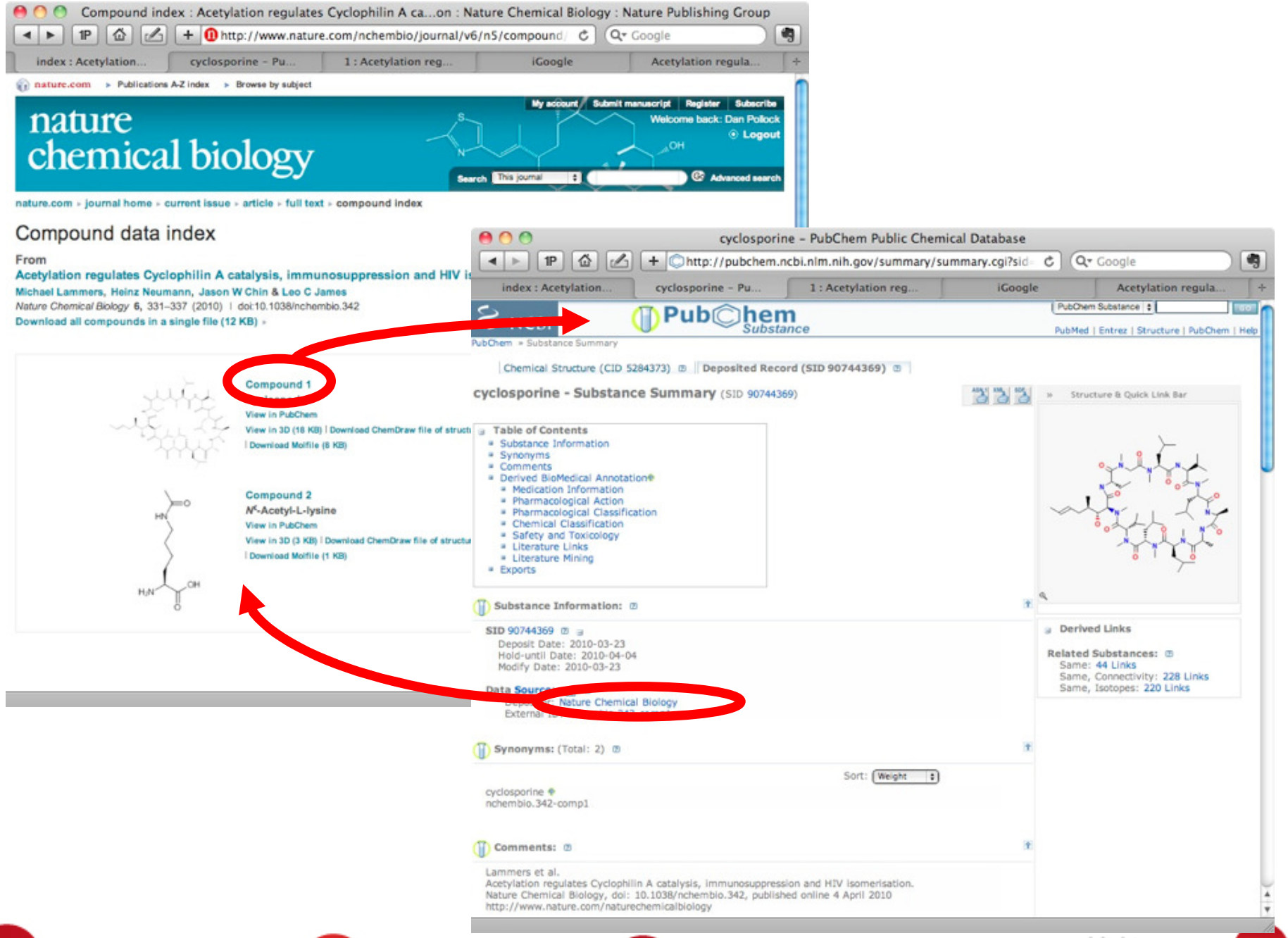
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Nature Chemical Biology 6, 331–337 (2010) | doi:10.1038/nchembio.342
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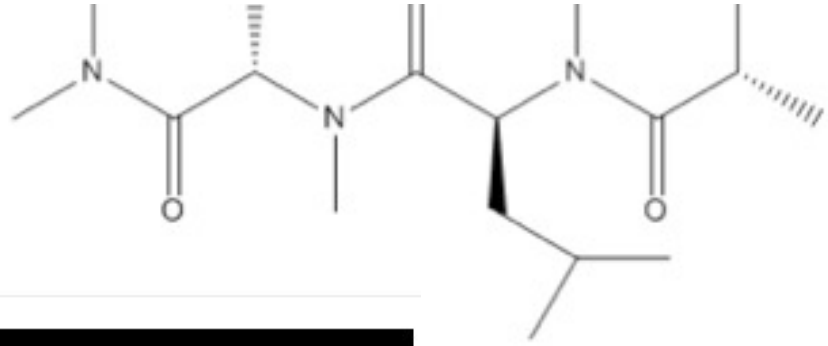
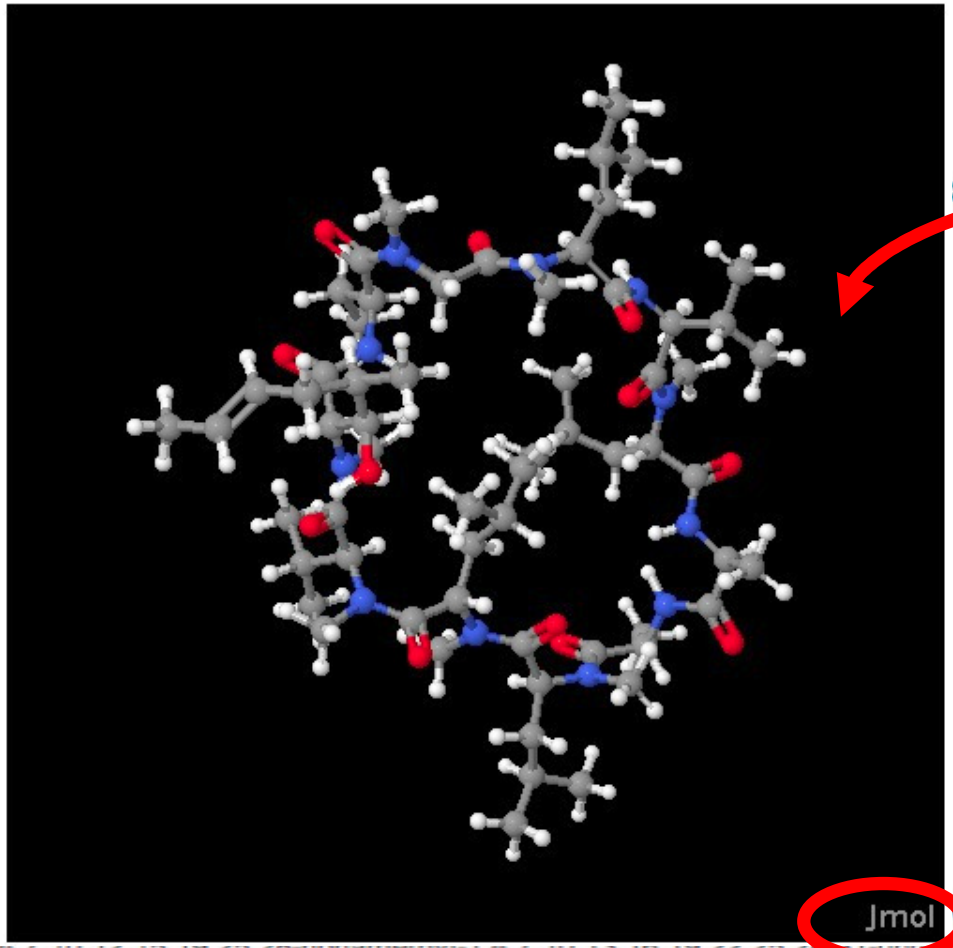


Compound 1: Cyclosporine



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caazacyclotritriacontane-2,5,8,11,14,17,20,23,26,29,32-undecone

1,4,7,10,13,16,19,22,25,28,31-undecaazacyclotritriacontane-2,5,8,11,14,17,20,23,26,29,32-undecone, 30-ethyl-33-[(1R,2R,4E)-1-hydroxy-2

1,4,7,10,12,15,19,25,28-nonamethyl-3,21-bis(1-m

ethylethyl)-6,9,18,24-tetrakis(2-methylpropyl)-, (3S,6S,9S,12R,15S,18S,21S,24S,30S,33S)-

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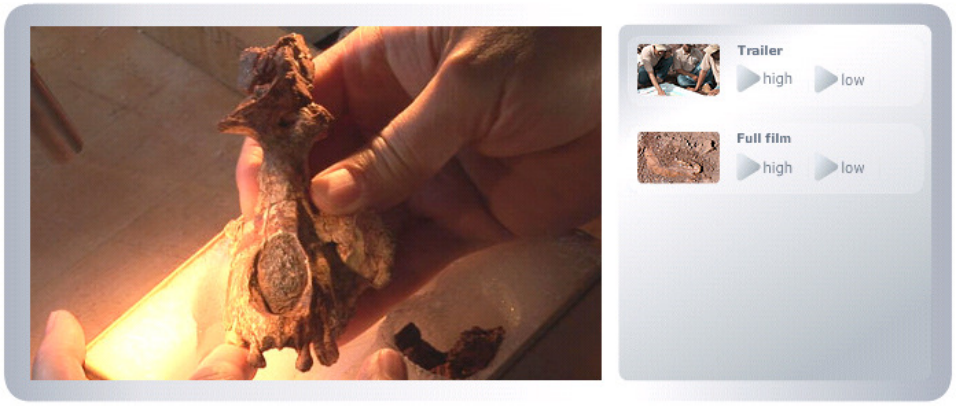

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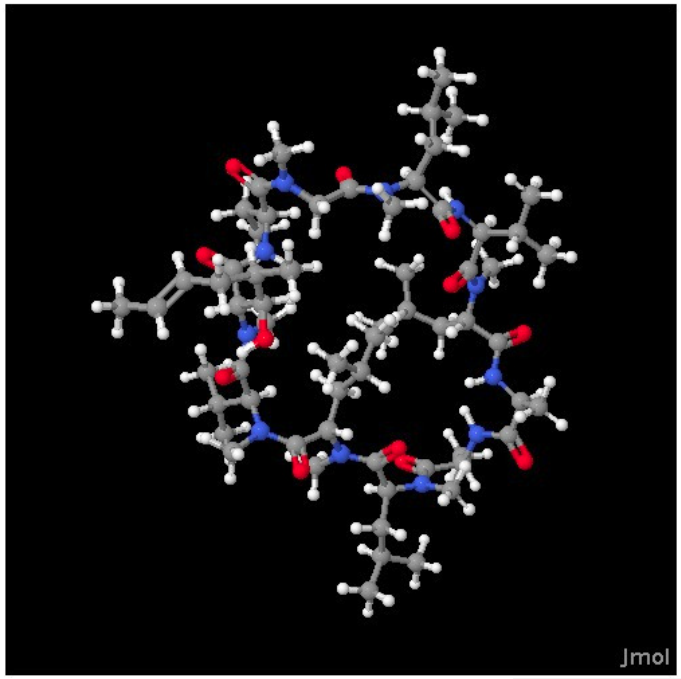

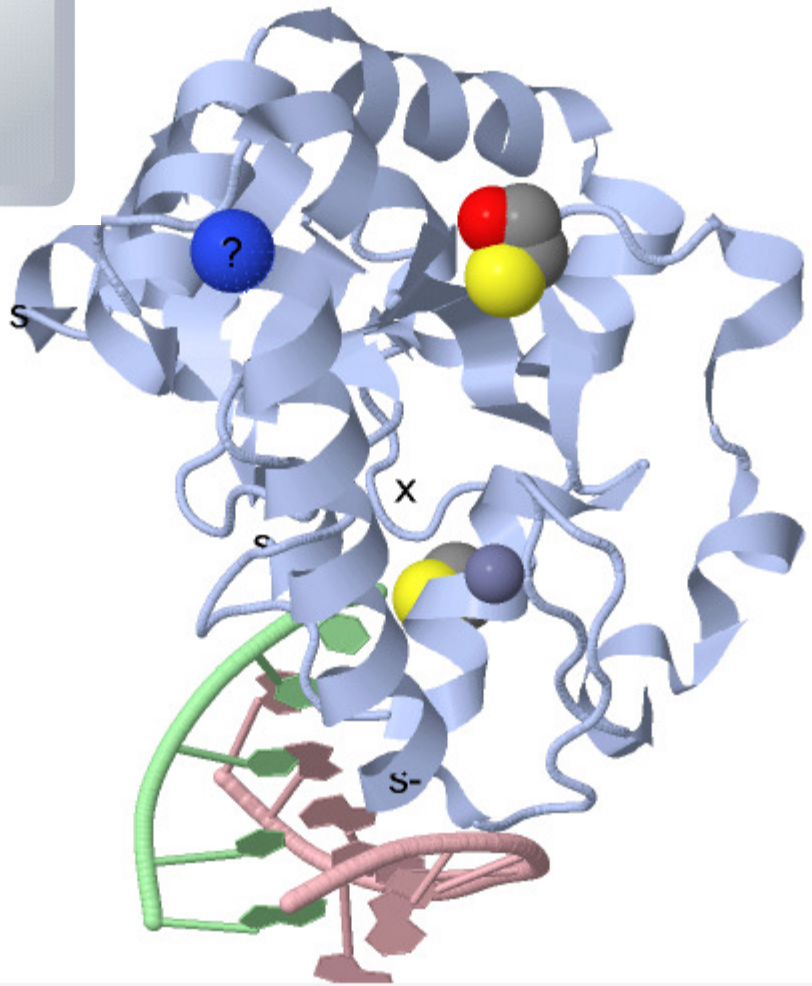

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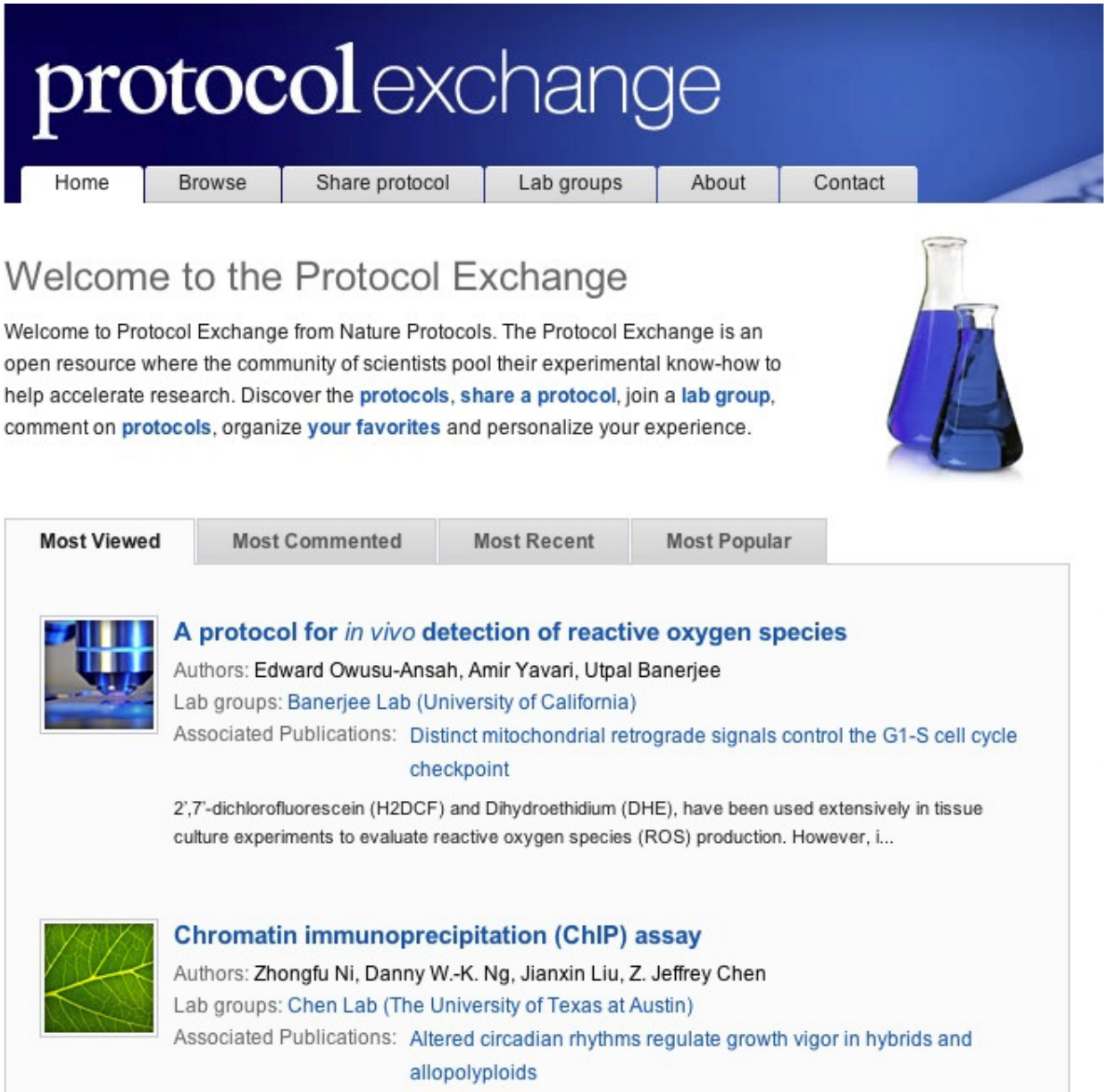

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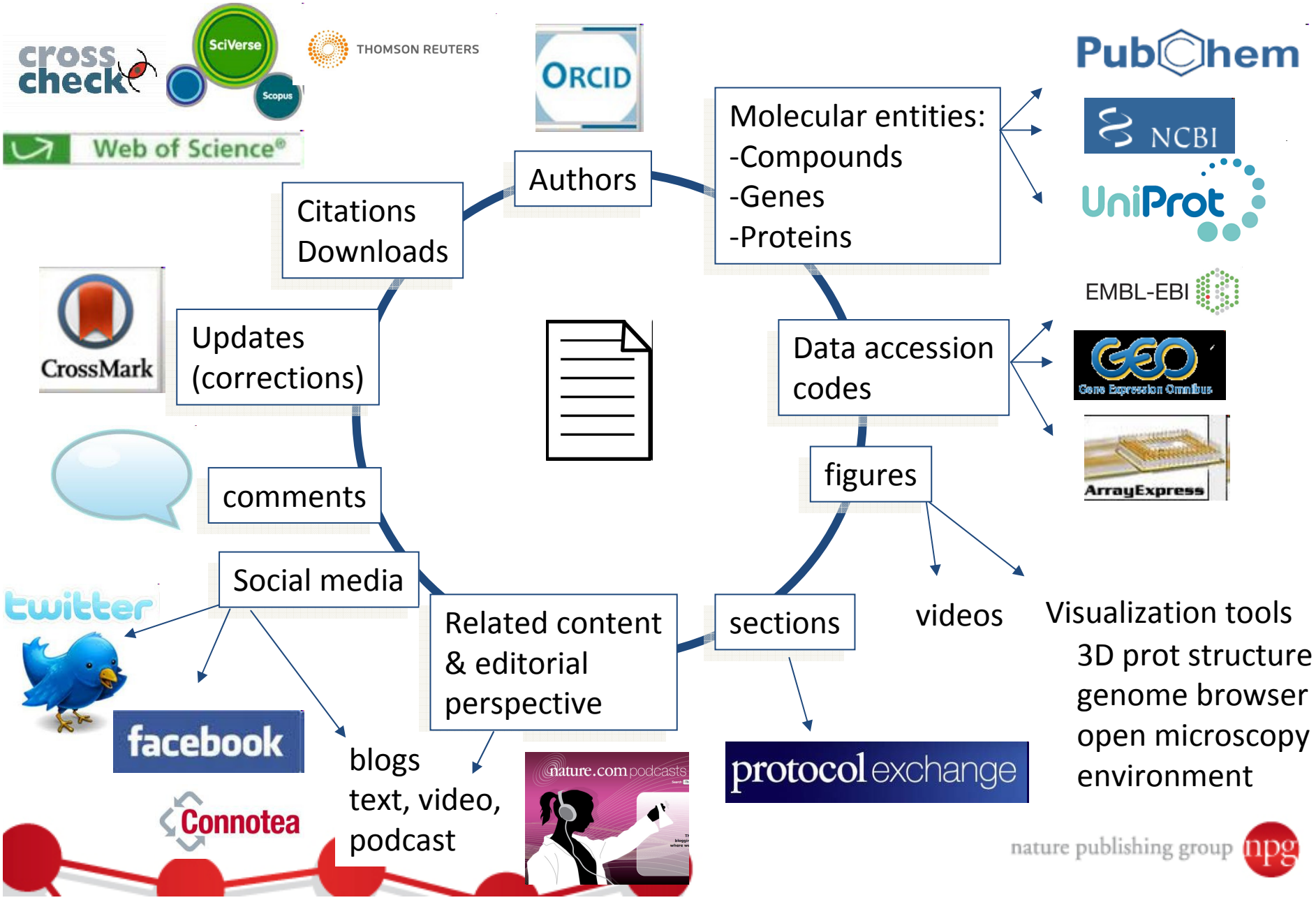
- Users join online groups, align their identify their Labs.

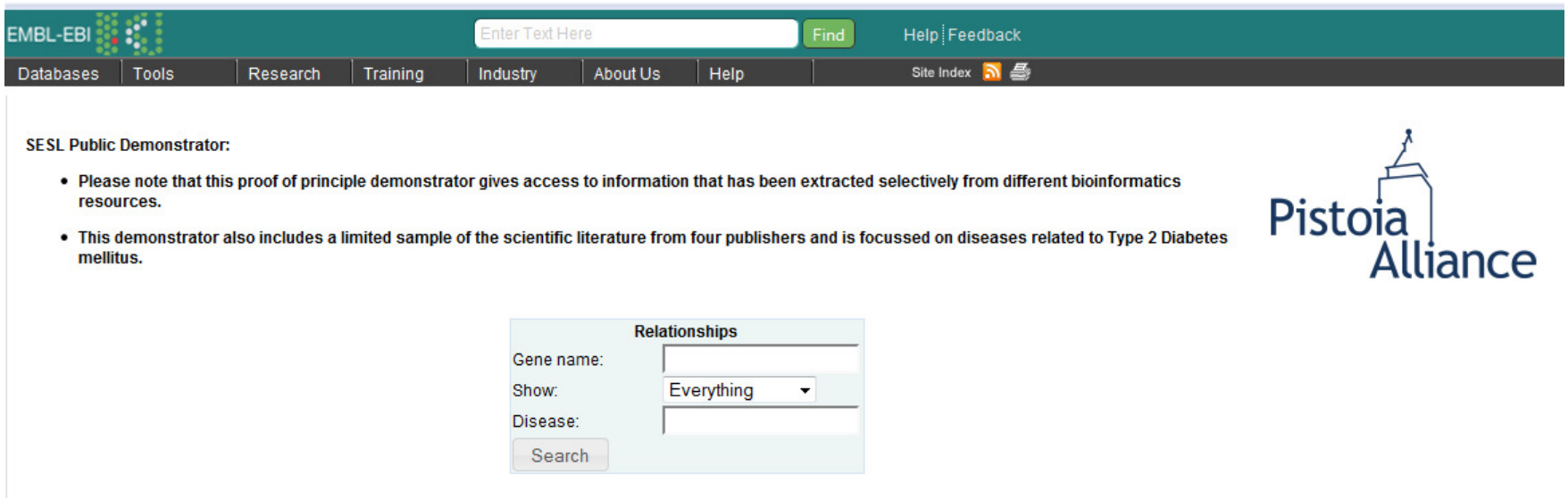
Atomising the unit of publication



Cross industry standards

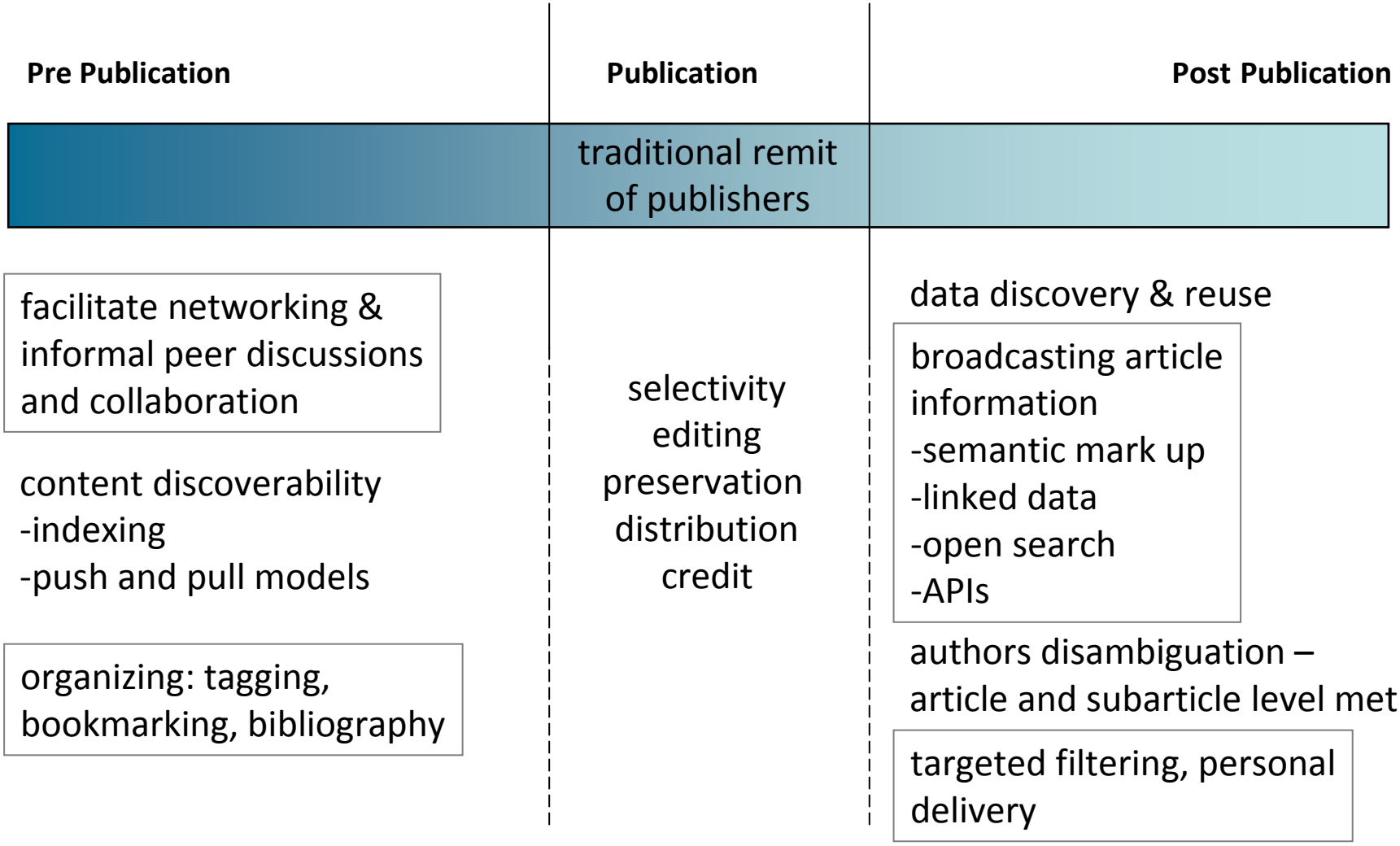
- Credit where credit is due
  - Disambiguation
    - doi, ORCID
  - Performance measures
    - ISI impact factor, COUNTER, PIRUS, SUSHI
  - Editorial quality control
    - crosscheck, crossmark
- Openness
  - Broadcasting in machine readable format
    - Opensearch, RDF triple stores, linked data
    - APIs and external developers programmes

Experimenting with push versus pull



- Pistoia SESL project
- Databases: Uniprot, OMIM, and ArrayExpress
- 5 publishers: Nature Publishing Group, Oxford University Press, the Royal Society of Chemistry, UK Pubmed Central, and Elsevier.

An expanding remit



More will happen…